

# From vitamin D to hormone D: fundamentals of the vitamin D endocrine system essential for good health<sup>1-4</sup>

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## ABSTRACT

New knowledge of the biological and clinical importance of the steroid hormone  $1\alpha,25$ -dihydroxyvitamin  $D_3$  [ $1\alpha,25(OH)_2D_3$ ] and its receptor, the vitamin D receptor (VDR), has resulted in significant contributions to good bone health. However, worldwide reports have highlighted a variety of vitamin D insufficiency and deficiency diseases. Despite many publications and scientific meetings reporting advances in vitamin D science, a disturbing realization is growing that the newer scientific and clinical knowledge is not being translated into better human health. Over the past several decades, the biological sphere of influence of vitamin  $D_3$ , as defined by the tissue distribution of the VDR, has broadened at least 9-fold from the target organs required for calcium homeostasis (intestine, bone, kidney, and parathyroid). Now, research has shown that the pluripotent steroid hormone  $1\alpha,25(OH)_2D_3$  initiates the physiologic responses of  $\geq 36$  cell types that possess the VDR. In addition to the kidney's endocrine production of circulating  $1\alpha,25(OH)_2D_3$ , researchers have found a paracrine production of this steroid hormone in  $\geq 10$  extrarenal organs. This article identifies the fundamentals of the vitamin D endocrine system, including its potential for contributions to good health in 5 physiologic arenas in which investigators have clearly documented new biological actions of  $1\alpha,25(OH)_2D_3$  through the VDR. As a consequence, the nutritional guidelines for vitamin  $D_3$  intake (defined by serum hydroxyvitamin  $D_3$  concentrations) should be reevaluated, taking into account the contributions to good health that all 36 VDR target organs can provide. *Am J Clin Nutr* 2008;88(suppl):491S-9S.

## INTRODUCTION

Vitamin  $D_3$  is essential for life in higher animals. Research has shown, for example, that vitamin  $D_3$  is one of the primary biological regulators of calcium homeostasis. Vitamin  $D_3$ 's important biological effects occur only as a consequence of its metabolism into a family of daughter metabolites, including the key kidney-produced metabolite  $1\alpha,25$ -dihydroxyvitamin  $D_3$  [ $1\alpha,25(OH)_2D_3$ ]. Researchers consider  $1\alpha,25(OH)_2D_3$  to be a steroid hormone and believe that it functions the same way as other steroid hormones—by interacting with its cognate vitamin D receptor (VDR) (1).

The role of vitamin  $D_3$  as a vitamin or essential dietary component, in concert with the biological and clinical importance of the steroid hormone  $1\alpha,25(OH)_2D_3$  and the VDR, has achieved increasing prominence over the past 3 to 4 decades in the public health arena because of its contributions to good health in the general public. However, despite the plethora of publications and scientific meetings focusing on advances in vitamin D science, there is the disturbing realization that all is not “well” with the translation of the newer

scientific and clinical knowledge into the achievement of better health. Scientists and nutrition experts at the 13th Vitamin D Workshop held in 2006 agreed in a consensus statement that “about half of the elderly in North America and two-thirds of the rest of the world are not getting enough vitamin D to maintain healthy bone density, lower their risks for fractures and improve tooth attachment. Such vitamin D insufficiency also decreases muscle strength and increases the risk for falls and is even associated with increased risk for colorectal and other major cancer” (2).

The purpose of this article is to identify the fundamentals of the vitamin D endocrine system and the actions of  $1\alpha,25(OH)_2D_3$  that are essential for good health, keeping in mind that the totality of their contributions depends on the adequate availability of 25-hydroxyvitamin  $D_3$  [ $25(OH)D_3$ ], which in turn depends on appropriate vitamin D nutritional status as determined by ultraviolet (UV) radiation exposure and dietary intake of vitamin  $D_3$ .

## SOURCES OF VITAMIN D

Vitamin D is not technically a vitamin, ie, it is not an essential dietary factor; rather, it is a prohormone produced photochemically in the skin from 7-dehydrocholesterol. The molecular structure of vitamin D is closely allied to that of classic steroid hormones (eg, estradiol, cortisol, and aldosterone) in that they have the same root cyclopentanoperhydrophenanthrene ring structure. Technically, vitamin D is a secosteroid because one of the rings of its cyclopentanoperhydrophenanthrene structure has a broken carbon-carbon bond; in vitamin D, this occurs in the 9,10 carbon-carbon bond of ring B (Figure 1). Given that fact as a starting point, the reader must have access to some of the details of the sunlight-mediated photochemical conversion of 7-dehydrocholesterol into vitamin  $D_3$ ; this information is provided in Figure 1.

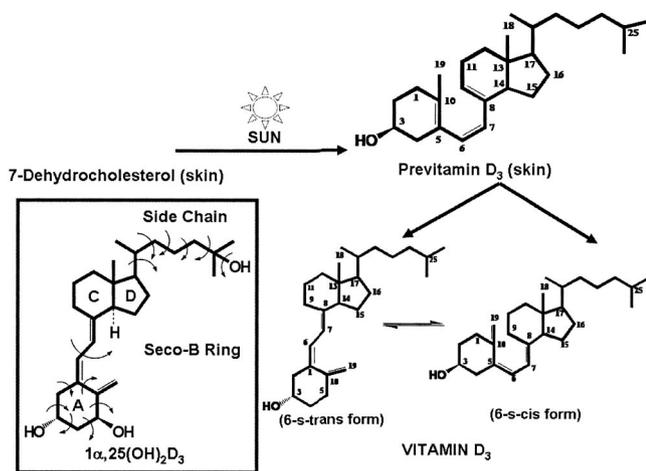
The skin produces vitamin  $D_3$  photochemically from the provitamin D, 7-dehydrocholesterol, which is present in the epidermis or skin of higher animals, by the action of sunlight in most geographical locations or of artificial UV light. The conjugated double-bond system in ring B (see Figure 1) allows the absorption of light quanta at

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**FIGURE 1.** Chemistry and irradiation pathway for the production of vitamin D<sub>3</sub>. The provitamin, which is characterized by the presence in the B ring of a  $\Delta 5, \Delta 7$  conjugated double bond system, is converted to a seco B provitamin steroid after exposure to ultraviolet light and the 9,10 carbon-carbon bond is broken. Next, in a process independent of ultraviolet light, the provitamin D thermally isomerizes to the vitamin form, which is characterized by a  $\Delta 6,7, \Delta 8,9, \Delta 10, 19$  conjugated triple bond system. The main portion of the figure also illustrates the 2 principal conformations of the molecule that result from rotation about the 6,7 carbon single bond of the seco B ring: the 6-*s-cis* conformer (the steroid-like shape) and the 6-*s-trans* conformer (the extended shape). The interconversion of the 2 conformers occurs millions of times per second. The extreme conformational flexibility potential of all vitamin D metabolites is illustrated in the inset for the principal metabolite 1 $\alpha,25$ -dihydroxyvitamin D<sub>3</sub> [1 $\alpha,25$ (OH)<sub>2</sub>D<sub>3</sub>]. Each arrow indicates carbon-carbon single bonds (in the side chain, the seco B ring, and the A ring) that have 360° rotational freedom. For all vitamin D molecules, this results in a multitude of different shapes in solution and in biological systems (8).

certain wavelengths in the UV range, initiating a complex series of transformations of the provitamin (partially summarized in Figure 1) that ultimately result in the generation of vitamin D<sub>3</sub>. Thus, vitamin D<sub>3</sub> can be endogenously produced. As long as the animal (or human) has access to adequate sunlight on a regular basis, the animal (or human) might not need to obtain this vitamin from the diet. However, dermatologists are concerned that individuals with extensive UV radiation exposure could have an increased risk of skin cancer or melanoma (3). A discussion of the parameters affecting the balance between sun exposure and dietary vitamin D to meet bodily requirements is available elsewhere (4). Because vitamin D<sub>3</sub> is also a vitamin, animals and humans can meet some or all of their vitamin D<sub>3</sub> needs through the diet.

For the past 6 decades, the literature has claimed that vitamin D<sub>3</sub> and vitamin D<sub>2</sub> (see Figure 2 for their structures) have equivalent biological effects in humans. However, with the realization that the serum 25(OH)D clinical assay provides the best assessment of vitamin D nutritional status (5), researchers needed to determine whether vitamin D<sub>2</sub> is as effective in elevating serum 25(OH)D concentrations in humans as is vitamin D<sub>3</sub>. Heaney and others recently confirmed earlier data suggesting that vitamin D<sub>3</sub> is substantially more effective than vitamin D<sub>2</sub> (6, 7). In a study of 20 healthy human volunteers, these investigators found that vitamin D<sub>2</sub>'s potency was less than one-third of vitamin D<sub>3</sub>'s potency on the basis of ability to elevate serum 25(OH)D concentrations.

## VITAMIN D ENDOCRINE SYSTEM

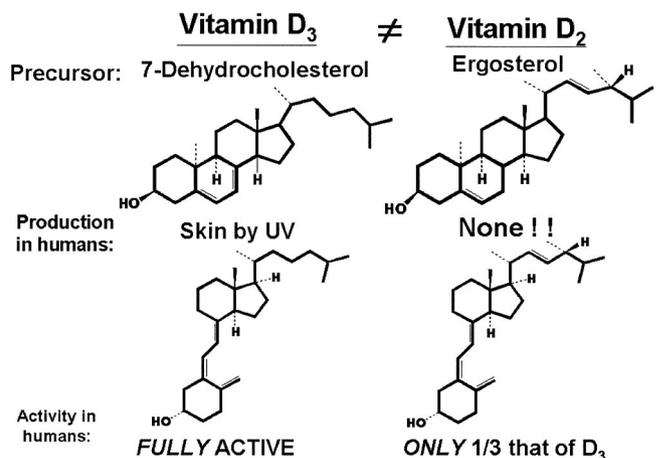
Vitamin D<sub>3</sub> does not have any known intrinsic biological activity. It must first be metabolized to 25(OH)D<sub>3</sub> in the liver and then to

1 $\alpha,25$ (OH)<sub>2</sub>D<sub>3</sub>, 24R,25-dihydroxyvitamin D<sub>3</sub> [24R,25(OH)<sub>2</sub>D<sub>3</sub>], or both by the kidney. Researchers have isolated and chemically characterized some 37 vitamin D<sub>3</sub> metabolites (8). Investigators have also established that humans and some other animals can metabolize vitamin D<sub>2</sub> to 25(OH)D<sub>2</sub> and 1 $\alpha,25$ (OH)<sub>2</sub>D<sub>2</sub> and many other similar cognate metabolites (9).

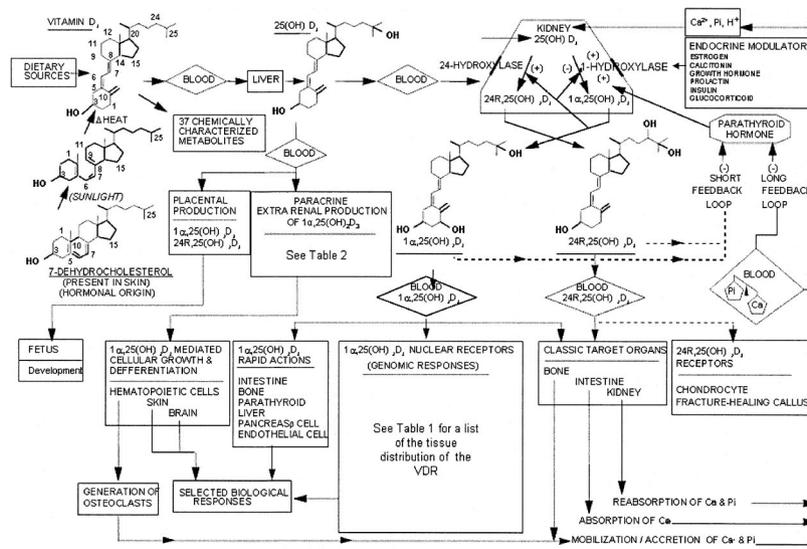
The steps in the vitamin D endocrine system (8) include the following (Figure 3): 1) the photoconversion of 7-dehydrocholesterol to vitamin D<sub>3</sub> in the skin or dietary intake of vitamin D<sub>3</sub>; 2) metabolism of vitamin D<sub>3</sub> by the liver to 25(OH)D<sub>3</sub>, the major form of vitamin D circulating in the blood compartment; 3) conversion of 25(OH)D<sub>3</sub> by the kidney (functioning as an endocrine gland) to the hormone 1 $\alpha,25$ (OH)<sub>2</sub>D<sub>3</sub> and the candidate hormone 24R,25(OH)<sub>2</sub>D<sub>3</sub>; 4) systemic transport of the dihydroxylated metabolites 24R,25(OH)<sub>2</sub>D<sub>3</sub> and 1 $\alpha,25$ (OH)<sub>2</sub>D<sub>3</sub> to distal target organs; and 5) binding of 1,25(OH)<sub>2</sub>D<sub>3</sub> to a nuclear receptor, plasma membrane receptor, or both at the target organs, followed by generation of appropriate biological responses. An additional key component in the operation of the vitamin D endocrine system is the plasma vitamin D binding protein, which carries vitamin D<sub>3</sub> and its metabolites to their metabolism and target organs (10).

The most important regulation point in the classic vitamin D endocrine system occurs through the stringent control of the circulating concentration of the steroid hormone 1 $\alpha,25$ (OH)<sub>2</sub>D<sub>3</sub>; typically, its production is modulated according to the organism's calcium and other endocrine needs. The chief regulatory factors are 1 $\alpha,25$ (OH)<sub>2</sub>D<sub>3</sub> itself, which down-regulates its own production; parathyroid hormone, which stimulates the renal production of 1,25(OH)<sub>2</sub>D<sub>3</sub>; fetal growth factor 23; and serum concentrations of calcium and phosphate (11). Probably the most important determinant of the 25(OH)D-1 $\alpha$ -hydroxylase activity is the animal's vitamin D nutritional status (11). When the circulating concentration of 1 $\alpha,25$ (OH)<sub>2</sub>D<sub>3</sub> is low, production of 1 $\alpha,25$ (OH)<sub>2</sub>D<sub>3</sub> by the kidney is high; when the circulating concentration of 1 $\alpha,25$ (OH)<sub>2</sub>D<sub>3</sub> is high, the output of 1 $\alpha,25$ (OH)<sub>2</sub>D<sub>3</sub> by the kidney decreases sharply (11).

The pervasive contributions of the VDR in collaboration with its ligand, 1 $\alpha,25$ (OH)<sub>2</sub>D<sub>3</sub>, to the functioning of the vitamin D endocrine system are illustrated in Figure 3 and listed in Table 1. Thirty-six tissues definitively possess the VDR, which means that the cells in these tissues have the potential to produce biological responses, depending on the availability of appropriate amounts of vitamin D<sub>3</sub>.



**FIGURE 2.** Structural and biological similarities and differences between vitamin D<sub>3</sub> and vitamin D<sub>2</sub>.



**FIGURE 3.** The vitamin D endocrine system. In this system, the biologically inactive vitamin D<sub>3</sub> is activated, first in the liver to produce 25-hydroxyvitamin D<sub>3</sub> [25(OH)D<sub>3</sub>], and the endocrine gland (the kidney) converts it to the hormones 1α,25-dihydroxyvitamin D<sub>3</sub> [1α,25(OH)<sub>2</sub>D<sub>3</sub>] and 24R,25-dihydroxyvitamin D<sub>3</sub> [24R,25(OH)<sub>2</sub>D<sub>3</sub>]. Researchers have identified ≥36 target organs, defined by the presence of the vitamin D receptor (VDR), which is the receptor for the steroid hormone 1α,25(OH)<sub>2</sub>D<sub>3</sub>; see Table 1 for a list of these target organs. Table 2 lists the 10 extrarenal tissues that investigators have shown possess the 1α-hydroxylase, the enzyme that converts 25(OH)D<sub>3</sub> to the steroid hormone 1α,25(OH)<sub>2</sub>D<sub>3</sub>. Pi, inorganic phosphate.

Barbour et al (12) discovered the first extrarenal 25(OH)D<sub>3</sub>-1α-hydroxylase in 1981 in a hypercalcemic anephric patient with sarcoidosis. Researchers now recognize that in individuals with any of a number of granulomatous diseases, the locally produced 1α,25(OH)<sub>2</sub>D<sub>3</sub> frequently spills over into the general circulation, which results in hypercalcemia. Researchers have shown that the enzyme that converts 25(OH)D<sub>3</sub> into 1α,25(OH)<sub>2</sub>D<sub>3</sub>, namely the 25(OH)D<sub>3</sub>-1α-hydroxylase, is present in a paracrine fashion in at least 10 tissues in addition to the proximal tubule of the kidney (Table 2). Thus, cells that express a functional 25(OH)D<sub>3</sub>-1α-hydroxylase acquire the ability to produce local concentrations of the steroid hormone 1α,25(OH)<sub>2</sub>D<sub>3</sub>. This “local” or modest

production of 1α,25(OH)<sub>2</sub>D<sub>3</sub> then generates biological responses in the local cellular neighborhood. It is believed that this paracrine-generated 1α,25(OH)<sub>2</sub>D<sub>3</sub> does not normally spill over into the circulatory system; thus, the plasma concentration of 1α,25(OH)<sub>2</sub>D<sub>3</sub> does not increase in a measurable way. The ability of locally produced 1α,25(OH)<sub>2</sub>D<sub>3</sub> to promote cell differentiation in prostate cancer (24) and colon cancer cells (25) are examples demonstrating its potential biological importance.

**TABLE 1**  
Tissues that express the vitamin D receptor for the steroid hormone 1α, 25-dihydroxyvitamin D<sub>3</sub><sup>1</sup>

Tissue distribution	
Adipose	Muscle, embryonic
Adrenal	Muscle, smooth
Bone	Osteoblast
Bone marrow	Ovary
Brain	Pancreas β cell
Breast	Parathyroid
Cancer cells	Parotid
Cartilage	Pituitary
Colon	Placenta
Eggshell gland	Prostate
Epididymis	Retina
Hair follicle	Skin
Intestine	Stomach
Kidney	Testis
Liver (fetal)	Thymus
Lung	Thyroid
Lymphocytes (B & T)	Uterus
Muscle, cardiac	Yolk sac (bird)

<sup>1</sup> For reference citations on the distribution of the vitamin D receptor, see reference 8.

**EXPANSION OF VITAMIN D KNOWLEDGE**

The consequences that the new knowledge of the vitamin D endocrine system and the various vitamin D metabolites has had on the rate of publication of peer-reviewed articles on vitamin D

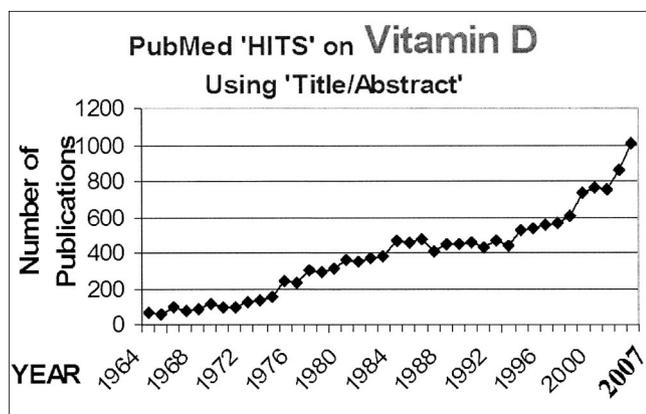
**TABLE 2**  
Sites of extrarenal 1α,25-dihydroxyvitamin D<sub>3</sub> production in humans<sup>1</sup>

Tissue	mRNA <sup>2</sup>	Protein <sup>2</sup>	Enzymatic activity <sup>3</sup>	Reference
Colon	Y	Y	Y	(13)
Dendritic cells	Y	—	—	(14)
Endothelial cells	Y	Y	Y	(15)
Human brain	—	Y	—	(16)
Mammary, breast	Y	Y	Y	(17)
Pancreatic islets	Y	Y	Y	(18)
Parathyroid glands	Y	Y	Y	(19)
Placenta, decidua	Y	Y	Y	(20)
Prostate	Y	Y	Y	(21)
Skin, keratinocytes	Y	Y	Y	(22, 23)

<sup>1</sup> Based on the presence of the 25-hydroxyvitamin D<sub>3</sub>-1α-hydroxylase enzyme. Hewison and colleagues have studied the tissue localization of the extrarenal 25-hydroxyvitamin D<sub>3</sub>-1α-hydroxylase enzyme extensively in humans (18, 20).

<sup>2</sup> Detection of mRNA for the 1α-hydroxylase and the 1α-hydroxylase protein.

<sup>3</sup> Presence of measurable enzymatic activity of the 25-hydroxyvitamin D<sub>3</sub>-1α-hydroxylase.



**FIGURE 4.** Growth in the number of articles published each year with the term *vitamin D* in the title or abstract, as reported in PUBMED (National Library of Medicine, Bethesda, MD).

is illustrated in **Figure 4**. In 1975, journals published only  $\approx 100$  articles per year that included the term *vitamin D* in the title or abstract; by 2007, the rate of publication had increased to  $>1400$  articles per year. Another driving factor for this increased publication rate was the chemical synthesis by academic chemists and by 4 pharmaceutical companies of  $>2000$  analogues of  $1\alpha,25(\text{OH})_2\text{D}_3$ ; most of these analogues were targeted toward selective responses in diseases such as osteoporosis, renal osteodystrophy, and psoriasis, and the authors reported their biological properties in a multitude of publications (8, 26).

Thus, an enormous body of scientific literature currently exists for vitamin D. PUBMED (National Library of Medicine, Bethesda, MD) lists  $>20\,700$  publications that use the term *vitamin D* in either the title or abstract from 1950 to the present. This total includes articles that combine the use of *vitamin D* with one of the following terms: *bone* ( $>6300$  articles), *deficiency* ( $>2900$ ), *cancer* ( $>1500$ ), *renal failure* ( $>700$ ), *intestine* ( $>700$ ), *cardiovascular/heart* ( $>600$ ), *diabetes* ( $>470$ ), *insulin* ( $>450$ ), or *brain* ( $>270$ ). Finally, PUBMED lists  $>4500$  publications with the term *vitamin D<sub>3</sub>* in the title or abstract,  $>660$  articles with *vitamin D<sub>2</sub>*, and  $>2600$  articles with *calcitriol* [a synonym for  $1\alpha,25(\text{OH})_2\text{D}_3$ ].

Unfortunately, the proliferation of published articles on vitamin D occurred at the same time as the use by many authors of less precise terminology to describe 3 key vitamin D molecules, namely, vitamin D<sub>3</sub>, vitamin D<sub>2</sub>, and  $1\alpha,25(\text{OH})_2\text{D}_3$ . Figure 2 emphasizes the structural differences between vitamins D<sub>3</sub> and D<sub>2</sub> and provides a clear description and comparison of the biological properties of these 2 key molecules. This figure shows that 1) vitamin D<sub>3</sub> is the only naturally occurring form of vitamin D in humans and other animals and 2) vitamin D<sub>2</sub> has only one-third the biological activity of vitamin D<sub>3</sub> in humans (7). Given the significant differences in the biological activity of vitamin D<sub>3</sub> and vitamin D<sub>2</sub>, describing a clinical trial by writing that “the patients received 10  $\mu\text{g}$  (400 IU) of vitamin D” is not a best practice when the patients actually received vitamin D<sub>2</sub>, whose biological activity is equivalent to only 3.25  $\mu\text{g}$  (130 IU) vitamin D<sub>3</sub>.

Another frequent error occurs when authors use *vitamin D* as a synonym for  $1\alpha,25(\text{OH})_2\text{D}_3$ , sometimes even in the methods sections of articles. The very significant structural and biological differences between  $1\alpha,25(\text{OH})_2\text{D}_3$  and vitamin D<sub>3</sub> are emphasized in **Figure 5**, which clearly shows why authors must not use *vitamin D* to refer to  $1\alpha,25(\text{OH})_2\text{D}_3$ . If, for example, a reader sees

the following statement in the results or discussion section of an article, “the animals or subjects received a standard vitamin D dose that would not cause hypercalcemia,” he or she would make a serious error of interpretation if he or she had not carefully read the methods section to learn that “all subjects received a dose of 1.5 micrograms of  $1\alpha,25(\text{OH})_2\text{D}_3$ .”

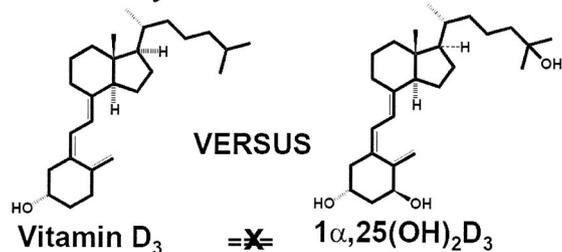
#### MODE OF ACTION OF $1\alpha,25(\text{OH})_2\text{D}_3$ : GENOMIC ACTIONS

The steroid hormone  $1\alpha,25(\text{OH})_2\text{D}_3$  and many other steroid hormones (eg, estradiol, progesterone, testosterone, cortisol, and aldosterone) generate biological responses both by regulating gene transcription (the classic genomic responses) and by rapidly activating a variety of signal transduction pathways at or near the plasma membrane (rapid or nongenotropic responses) (27). The genomic responses to  $1\alpha,25(\text{OH})_2\text{D}_3$  result from its stereospecific interaction with its nuclear receptor,  $\text{VDR}_{\text{nuc}}$  (**Figure 6**). The  $\text{VDR}_{\text{nuc}}$  is a protein of 50 kDa, which binds  $1\alpha,25(\text{OH})_2\text{D}_3$  with high affinity ( $K_d \approx 0.5$  nmol/L). The  $\text{VDR}_{\text{nuc}}$  does not bind the parent vitamin D<sub>3</sub> or vitamin D<sub>2</sub>;  $25(\text{OH})\text{D}_3$  and  $1\alpha(\text{OH})\text{D}_3$  only bind 0.1–0.3% as well as  $1\alpha,25(\text{OH})_2\text{D}_3$ . As is true for all nuclear receptors for the steroid hormones involved, the primary amino acid sequence of the  $\text{VDR}_{\text{nuc}}$  consists of 6 functional domains: the variable regions (A and B domains), DNA binding (the C domain), the hinge region (D domain), the ligand-binding region (E domain), and transcriptional activation (domain F) (28). A detailed discussion of the  $\text{VDR}_{\text{nuc}}$  and its participation in the regulation of gene transcription is available elsewhere (28).

Nuclear-receptor-mediated regulation of gene transcription depends on the exquisite structural relation between the unoccupied receptor, which is transcriptionally inactive, and its cognate ligand  $1\alpha,25(\text{OH})_2\text{D}_3$ . Formation of the ligand-receptor complex, which results in conformational changes in the receptor protein, then allows the ligand-receptor complex to specifically interact with the many proteins that collectively constitute the transcriptional machinery. The complementarity of the ligand shape with that of the interior surface of the nuclear VDR ligand binding domain is key not only to the structural basis of receptor action and its formation of heterodimers and interactions with coactivators (**Figure 7**), but also to designing new drug forms of

#### Vitamin D<sub>3</sub> vs $1\alpha,25(\text{OH})_2\text{-Vitamin D}_3$

Are they the same or different?



Properties:

Biologically Inactive Itself

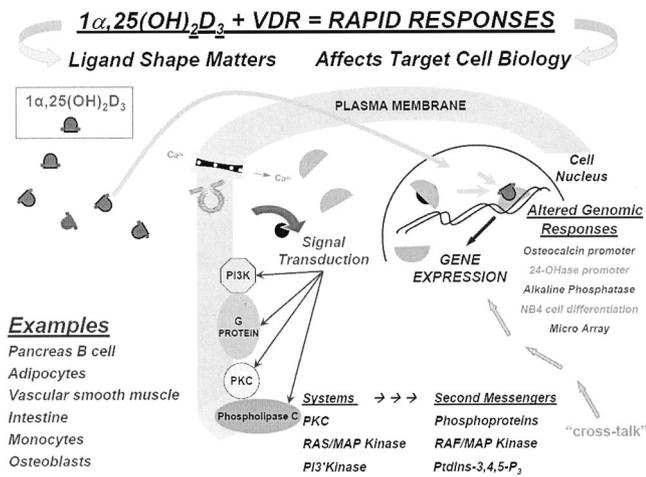
Steroid Hormone

Does not bind to VDR

Acts through VDR

Nutritional Substance

**FIGURE 5.** Structural and biological similarities and differences between vitamin D<sub>3</sub> and  $1\alpha,25(\text{OH})_2\text{D}_3$ . VDR, vitamin D receptor.



**FIGURE 6.** Schematic model showing how the conformationally flexible 1,25-dihydroxyvitamin D<sub>3</sub> [1,25(OH)<sub>2</sub>D<sub>3</sub>] can interact with the vitamin D receptor (VDR) localized in the cell nucleus to generate genomic responses and in caveolae with the plasma membrane VDR to generate rapid responses. Binding of 1,25(OH)<sub>2</sub>D<sub>3</sub> to the caveolae-associated VDR may result in the activation of one or more second messenger systems, including phospholipase C (PKC), protein kinase C, G protein-coupled receptors, or phosphatidylinositol-3-kinase (PI3K). The many possible outcomes include opening the voltage-gated calcium or chloride channels or generating the indicated second messengers. Some of these second messengers, particularly RAF/MAPK, can engage in crosstalk with the nucleus to modulate gene expression. PtdIns-3,4,5-P<sub>3</sub>, phosphatidylinositol-3,4,5-trisphosphate.

the various hormones, including 1,25(OH)<sub>2</sub>D<sub>3</sub>. Researchers estimate that the VDR can regulate the expression of as many as 500 of the ≈20 488 genes in the human genome (29). The large number of VDR-regulated genes undoubtedly reflects the consequence of the distribution of both the VDR and 25(OH)D<sub>3</sub>-1α-hydroxylase to many organs.

**MODE OF ACTION OF 1,25(OH)<sub>2</sub>D<sub>3</sub>: RAPID RESPONSES**

Investigators originally postulated that the “rapid” or nongenomic responses mediated by 1,25(OH)<sub>2</sub>D<sub>3</sub> were mediated through the interaction of the secosteroid with a novel protein receptor located on the cell’s external membrane. Researchers have shown more recently that this membrane receptor is the classic VDR (previously found primarily in the nucleus and cytosol) associated with caveolae present in the plasma membrane of a variety of cells (30). Caveolae are flask-shaped membrane invaginations enriched in sphingolipids and cholesterol that are commonly found in a wide variety of cells (31). Using VDR knockout and wild-type mice, researchers found that rapid modulation of osteoblast ion channel responses by 1,25(OH)<sub>2</sub>D<sub>3</sub> require the presence of a functional vitamin D nuclear and caveolae VDR receptor (32, 33).

Careful research using a variety of structural analogues of 1,25(OH)<sub>2</sub>D<sub>3</sub> has shown that the genomic and nongenomic responses to this conformationally flexible steroid hormone have different requirements for ligand structure (34). For example, a key consideration is the position of rotation about the 6,7 single carbon-carbon bond that can be either the 6-*s-cis* or 6-*s-trans* orientation (see Figure 1). The preferred shape of the ligand for VDR<sub>nuc</sub>, determined from the X-ray crystal structure of the receptor occupied with the ligand 1,25(OH)<sub>2</sub>D<sub>3</sub>, is a 6-*s-trans* shaped bowl with the A-ring 30° above the plane of the C/D rings. In contrast, structure-function

studies of rapid nongenomic actions of 1,25(OH)<sub>2</sub>D<sub>3</sub> and its analogues show that the VDR<sub>mem</sub> preferentially binds a ligand with a 6-*s-cis* shape (27). This new ligand structure-function knowledge will allow chemists to synthesize analogues of 1,25(OH)<sub>2</sub>D<sub>3</sub> that are selective for either genomic or rapid responses, depending on the ligand’s overall shape.

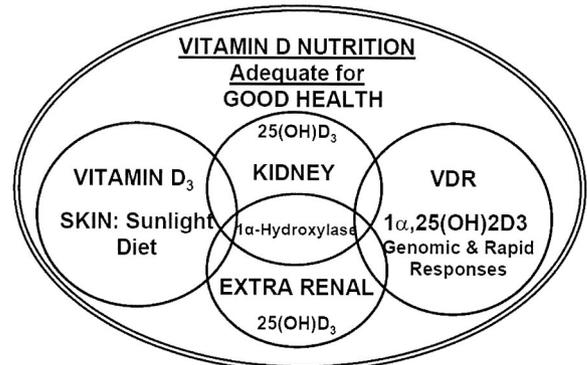
**DETERMINATION OF VITAMIN D STATUS**

As stated in the Introduction, the actions of 1,25(OH)<sub>2</sub>D<sub>3</sub> essential for good health depend on the vitamin D endocrine system. The operation of the vitamin D endocrine system to produce the steroid hormone 1,25(OH)<sub>2</sub>D<sub>3</sub> depends on the circulating concentration of 25(OH)D<sub>3</sub>; this key metabolite is the substrate for the 25(OH)D<sub>3</sub>-1α-hydroxylase enzyme that produces 1,25(OH)<sub>2</sub>D<sub>3</sub>. As is the case for any enzyme, the activity of the 1α-hydroxylase depends on the absolute concentration of its substrate. The K<sub>m</sub> or substrate concentration of 25(OH)D<sub>3</sub> required for 50% maximal activity for the 1α-hydroxylase is ≈100 nmol/L (11). As emphasized in Figure 2, the availability of 25(OH)D<sub>3</sub> depends on adequate access to vitamin D<sub>3</sub>. Thus, determining vitamin D nutritional status becomes a critical issue in optimizing the prospects for those aspects of “good health” that 1,25(OH)<sub>2</sub>D<sub>3</sub> can mediate or to which it can contribute.

Surprisingly, despite extensive efforts, no routine clinical assay is available for determining the serum concentration of either vitamin D<sub>3</sub> or vitamin D<sub>2</sub>. Furthermore, researchers are unlikely to develop a routine serum vitamin D clinical assay in the future. However, the US Institute of Medicine has endorsed the view that the circulating concentration of 25(OH)D<sub>3</sub> is an acceptable functional measure of vitamin D nutritional status (5, 35).

Information on obtaining insight into vitamin D nutritional status by determining the serum concentrations of several vitamin D metabolites is presented in **Tables 3** and **4**. Table 34 presents a tabulation of the circulating serum concentrations of the 3 major vitamin D metabolites: 25(OH)D<sub>3</sub>, 24,25(OH)D<sub>3</sub>, and 1,25(OH)<sub>2</sub>D<sub>3</sub>. The molar ratio of the total (not free steroid) serum concentrations of these metabolites is 830:77:1 for 25(OH)D to 24,25(OH)D<sub>3</sub> to 1,25(OH)<sub>2</sub>D<sub>3</sub>. Investigators primarily measure circulating concentrations of 24,25(OH)D<sub>3</sub> for

**PLANNING FOR VITAMIN D AND HEALTH IN 21<sup>ST</sup> CENTURY**



**MUST CONSIDER THE REALITY OF THE VITAMIN D ENDOCRINE SYSTEM**

**FIGURE 7.** Fundamentals of the vitamin D endocrine system that contribute to the daily maintenance of a vitamin D nutritional status that is essential for achieving good health. 25(OH)D<sub>3</sub>, 25-hydroxyvitamin D<sub>3</sub>; 1,25(OH)<sub>2</sub>D<sub>3</sub>, 1,25-dihydroxyvitamin D<sub>3</sub>.

experimental animal studies or selected clinical studies; such measurements are not available through commercial laboratories. Many, but not all, clinical chemistry laboratories can measure  $1\alpha,25(\text{OH})_2\text{D}_3$  concentrations. Because serum  $1\alpha,25(\text{OH})_2\text{D}_3$  values do not correlate with clinical disease status, information on serum  $1\alpha,25(\text{OH})_2\text{D}_3$  concentration does not usually help with clinical diagnosis and treatment (48).

Most researchers agree that the range of the serum concentration of  $25(\text{OH})\text{D}_3$  in a population of healthy subjects is the best indicator for assessing the vitamin D status in patients with a vitamin D-related disease. The factors supporting this include: 1) no vitamin D clinical assay is available; 2) the metabolism of vitamin  $\text{D}_3$  into  $25(\text{OH})\text{D}_3$  by the liver vitamin D-25-hydroxylase is not regulated, so the serum concentration of  $25(\text{OH})\text{D}_3$  is an accurate "reporter" of both cutaneous UV-stimulated synthesis and dietary intake of vitamin  $\text{D}_3$ ; 3) a variety of clinical assays are available to measure  $25(\text{OH})\text{D}$ ; and 4) the plasma concentrations of  $25(\text{OH})\text{D}_3$  correlate with many clinical diseases (6, 46). Hollis et al (41) argued that the relation between vitamin  $\text{D}_3$  and  $25(\text{OH})\text{D}_3$  is *not* linear, but rather saturable and controlled. They conclude that "optimal vitamin D status was achieved when  $25(\text{OH})\text{D}_3$  was  $> 40 \text{ ng/mL}$  or  $> 100 \text{ nmol/L}$ ," which is approximately equivalent to the  $K_m$  of the  $25(\text{OH})\text{D}_3$ - $1\alpha$ -hydroxylase.

Presented in Table 4 is a classification of circulating levels of  $25(\text{OH})\text{D}$  in relation to vitamin D nutritional status that was largely obtained from clinical studies relating to calcium homeostasis (intestinal calcium absorption, bone mineral density, parathyroid hormone concentrations, etc). In a large population of vitamin D-replete subjects, the normal range of  $25(\text{OH})\text{D}$  was found to be 25–137 nmol/L. But it was also noted that the lower limit of the normal range can vary among populations, ranging from as low as 20 up to 50 nmol/L (5). Undoubtedly, it will ultimately be essential to determine the normal  $25(\text{OH})\text{D}$  range in all ethnic groups and geographical populations of the world (at all latitudes to reflect differing UV exposures). I believe that a major goal for the vitamin D field is to agree on the "normal  $25(\text{OH})\text{D}$  serum levels" that support all 36 VDR-containing target organs in all the world's population groups.

## FUNDAMENTALS OF VITAMIN D AND ITS ENDOCRINE SYSTEM FOR GOOD HEALTH

The purpose of the Adequate Intake recommendations for vitamin D put forth by the Food and Nutrition Board of the

**TABLE 3**  
Serum circulating concentrations of key vitamin D metabolites<sup>1</sup>

Vitamin D metabolite	Vitamin D concentration	Reference
Vitamin D	Not routinely measured <sup>2</sup>	—
$25(\text{OH})\text{D}$	50–100 nmol/L (20–40 ng/ml)	(36, 41)
$24\text{R},25(\text{OH})_2\text{D}$	5–12 nmol/L (2–5 ng/ml)	(37, 38)
$1\alpha,25(\text{OH})_2\text{D}$	50–125 pmol/L (20–50 pg/ml)	(39)

<sup>1</sup>  $25(\text{OH})\text{D}$ , 25-hydroxyvitamin D;  $24\text{R},25(\text{OH})_2\text{D}$ ,  $24\text{R},25$ -dihydroxyvitamin D;  $1\alpha,25(\text{OH})_2\text{D}$ ,  $1\alpha,25$ -dihydroxyvitamin  $\text{D}_3$ . Vitamin D (both  $\text{D}_2$  and  $\text{D}_3$ ) is quite difficult to measure because of its hydrophobicity, so vitamin D measurements require extensive HPLC (40).

A US Institute of Medicine report has endorsed the view that the circulating concentration of  $25(\text{OH})\text{D}$  is a functional measure of vitamin D nutritional status (5); see also reference 36.

**TABLE 4**  
Circulating concentrations of 25-hydroxyvitamin D [ $25(\text{OH})\text{D}$ ] by vitamin D nutritional status<sup>1</sup>

Serum $25(\text{OH})\text{D}$ range <sup>2</sup>	Vitamin D nutritional status	Reference
$>75 \text{ nmol/L}$ ( $>30 \text{ ng/mL}$ )	Sufficiency	(42)
$>50 \text{ nmol/L}$ ( $>20 \text{ ng/mL}$ )	Sufficiency	(2, 43)
$30\text{--}50 \text{ nmol/L}$ ( $12\text{--}20 \text{ ng/mL}$ )	Insufficiency	(44)
$12\text{--}30 \text{ nmol/L}$ ( $5\text{--}12 \text{ ng/mL}$ )	Deficiency	(43)
$<12 \text{ nmol/L}$ ( $<5 \text{ ng/mL}$ ) <sup>2</sup>	Severe deficiency	(43)

<sup>1</sup> The classification of  $25(\text{OH})\text{D}$  concentrations into sufficiency, insufficiency, deficiency, and severe deficiency represents the author's interpretation of definitions in the publications listed. Researchers have suggested 2 distinct minimum serum concentrations of  $25(\text{OH})\text{D}$  for vitamin D sufficiency:  $>50 \text{ nmol/L}$  and  $>75 \text{ nmol/L}$ . The serum concentrations listed in the table refer to the sums of the concentrations of  $25(\text{OH})\text{D}_3$  and  $25(\text{OH})\text{D}_2$ . Certain methods for measuring  $25(\text{OH})\text{D}$  concentration yield information on both  $25(\text{OH})\text{D}_3$  and  $25(\text{OH})\text{D}_2$  without distinguishing between the two. However, mass spectrometry provides discrete values for each form of  $25(\text{OH})\text{D}$ . The use of liquid chromatography–tandem mass spectrometry has made it possible to simultaneously and routinely determine the amount of  $25(\text{OH})\text{D}_3$  and  $25(\text{OH})\text{D}_2$  in a small blood sample (45–47); the major drawback is that this costs more than \$100 per determination. In addition, many physicians cannot interpret differences between  $25(\text{OH})\text{D}_3$  and  $25(\text{OH})\text{D}_2$  concentrations.

<sup>2</sup> Persons with a  $25(\text{OH})\text{D}_3$  concentration  $<20 \text{ nmol/L}$  probably have rickets or osteomalacia (5).

Institute of Medicine (5) in 1999 was to provide guidelines of vitamin  $\text{D}_3$  intake to achieve normal serum levels of  $25(\text{OH})\text{D}$ . This was a very difficult goal to achieve, however, given that a quantitative relation of vitamin D's (ie, operation of the vitamin D endocrine system) contribution to good health was not clearly appreciated by 1997.

**Tables 5 and 6** focus on the new biological actions of the steroid hormone  $1\alpha,25(\text{OH})_2\text{D}_3$  that must be carefully studied to appreciate their dependency on an adequate availability of vitamin  $\text{D}_3$  to generate biological responses that are mediated by  $1\alpha,25(\text{OH})_2\text{D}_3$  to be *fully* compatible with proper health for each individual in the population. The 2 historical roles of vitamin D, namely stimulation of intestinal calcium absorption and increasing the mineral content and the remodeling of bone, are summarized in Table 5. For each process (intestine or bone), historical reference citations are provided to create the foundation that vitamin D is crucial to bone mineral content and intestinal calcium absorption. Thus, between 1922 and 1924, the pioneers Mellanby (54), McCollum (55), and Goldblatt (56) made the separate bone-related discoveries, respectively, that 1) the treatment or prevention of rickets could be mediated by cod liver oil; 2) by feeding a new vitamin, termed vitamin D; or 3) by exposure of skin to UV irradiation. Then in 1937, Nicolayson (49) showed the potent actions of vitamin  $\text{D}_3$  on stimulating intestinal calcium absorption in rats. Also included in Table 5 are comparable modern observations implicating the participation of the VDR and  $1\alpha,25(\text{OH})_2\text{D}_3$  in both intestinal calcium absorption and bone remodeling. It is these actions of  $1\alpha,25(\text{OH})_2\text{D}_3$  and its VDR that were largely addressed in 1999 by the Food and Nutrition Boards' guidelines for Adequate Intake of vitamin  $\text{D}_3$ .

Summarized in Table 6 are the 5 physiologic arenas in which new biological actions of  $1\alpha,25(\text{OH})_2\text{D}_3$  and the VDR have been

**TABLE 5**Roles of vitamin D identified in the early 20th century<sup>1</sup>

VDR-dependent system	VDR present or involved	Observation	Key reference
Intestinal calcium absorption	Not yet discovered	Vitamin D stimulates intestinal calcium and phosphorus absorption.	(49)
Intestinal calcium absorption	Yes	1 $\alpha$ ,25(OH) <sub>2</sub> D <sub>3</sub> stimulates intestinal calcium absorption in chicks and humans.	(50–52)
Bone formation and resorption (remodeling)	Not yet discovered	Giving the patient a new dietary supplement, called vitamin D, or exposing the skin to ultraviolet irradiation can prevent or treat rickets.	(54–56)
Bone formation and resorption (remodeling)	VDR	The actions of 1 $\alpha$ ,25(OH) <sub>2</sub> D <sub>3</sub> on the osteoblast (bone formation) and crosstalk with the osteoclast result in bone resorption and overall bone remodeling.	(53)

<sup>1</sup> VDR, vitamin D receptor; 1 $\alpha$ ,25(OH)<sub>2</sub>D, 1 $\alpha$ ,25-dihydroxyvitamin D<sub>3</sub>.

clearly documented and which have the capability of contributing to better health through improved vitamin D nutritional status. These include understanding at the molecular level the involvement of the VDR and 1 $\alpha$ ,25(OH)<sub>2</sub>D<sub>3</sub> (in both genomic and rapid responses) in 1) the functioning of B and T lymphocytes, which constitutes the adaptive immune system; 2) the innate immune system; 3) secretion of insulin by the pancreatic  $\beta$  cell; 4) the multifactorial functioning of the heart and blood pressure regulation; and 5) the multitude of activities of the brain. There are also multiple opportunities to address how the actions of the VDR and 1 $\alpha$ ,25(OH)<sub>2</sub>D<sub>3</sub> can affect several diseases. These include prevention and treatment of several cancers (breast, colon, and prostate) through prodifferentiation, antiproliferation, or induction of apoptosis effects, hypertension (infant heart failure), immunomodulation (psoriasis, type 1 diabetes, inflammatory bowel disease, periodontal disease, multiple sclerosis, and possibly rheumatoid arthritis), and neuromuscular effects (muscle

strength and better balance). Another critical goal is to understand the role of proper vitamin D nutrition and the molecular actions of the VDR with 1 $\alpha$ ,25(OH)<sub>2</sub>D<sub>3</sub> for all the steps of fetal development.

The important points made in this presentation are summarized in Figure 7. Researchers must understand the fundamentals of the vitamin D endocrine system, which include 1) vitamin D nutritional status (dietary intake and sunlight exposure), which define the circulating concentration of 25(OH)D<sub>3</sub> that is available to 2) the kidney functioning as an endocrine gland, which is complimented by 3) the paracrine 1 $\alpha$ -hydroxylase activities in 10 tissues to produce the steroid hormone 1 $\alpha$ ,25(OH)<sub>2</sub>D<sub>3</sub>, which will 4) interact with the VDR in  $\geq 36$  target tissues to generate optimal responses in the calcium homeostasis system. Researchers must also understand the 5 new physiologic systems that collectively comprise the vitamin D endocrine system.

**TABLE 6**Newly identified biological actions of 1 $\alpha$ , 25(OH)<sub>2</sub>D<sub>3</sub> with relevance to vitamin D nutritional status and resulting good health<sup>1</sup>

VDR-dependent system	VDR present or involved	Observation	Key reference
B and T lymphocytes	Yes	VDR is present in activated human mononuclear leukocytes and lymphocytes.	(57, 58)
Adaptive immune system	Yes	1,25-dihydroxyvitamin D <sub>3</sub> plays an immunoregulatory role.	(59)
Innate immune system	Yes	1 $\alpha$ ,25(OH) <sub>2</sub> D <sub>3</sub> induces cathelicidin, an antimicrobial peptide, through VDR-mediated gene expression.	(60)
Innate immune system	Yes	Toll-like receptor activation of human macrophages upregulates expression of the VDR and the 25(OH)D <sub>3</sub> -1 $\alpha$ -hydroxylase genes, leading to induction of the antimicrobial peptide cathelicidin and killing of intracellular <i>Mycobacterium tuberculosis</i> .	(61)
Innate immune system	Yes	Skin injury enhances antimicrobial peptide synthesis through the VDR and the 25(OH)D <sub>3</sub> -1 $\alpha$ -hydroxylase in keratinocytes.	(62)
Pancreas $\beta$ cells	Yes	Vitamin D deficiency inhibits pancreatic secretion of insulin and 1 $\alpha$ ,25(OH) <sub>2</sub> D <sub>3</sub> restores it.	(63, 64)
Pancreas $\beta$ cells	Yes	Vitamin D deficiency in early life accelerates development of type 1 diabetes in nonobese diabetic mice.	(65, 66)
Pancreas $\beta$ cells	Yes	In humans, 25(OH)D concentration has a positive correlation with insulin sensitivity and hypovitaminosis D has a negative effect on $\beta$ cell function.	(67)
Brain	Yes	The VDR and 1 $\alpha$ -hydroxylase are distributed in human brain.	(16)
Brain	Yes	Vitamin D deficiency in utero alters adult behavior in mice. Researchers have suggested that in humans, fetal deprivation of vitamin D <sub>3</sub> could be associated with adverse neuropsychiatric outcomes.	(68)
Brain	Yes	Combined prenatal and chronic postnatal vitamin D deficiency in rats impairs prepulse inhibition of acoustic startle.	(69)
Heart function and blood pressure regulation	Yes	Research characterized heart size and blood pressure in the VDR knockout mouse.	(70)
	Yes	Hypocalcaemia and vitamin D deficiency is an important but preventable cause of life-threatening infant heart failure.	(71)
	Yes	1 $\alpha$ ,25(OH) <sub>2</sub> D <sub>3</sub> is a negative endocrine regulator of the renin-angiotensin system and blood pressure.	(72)

<sup>1</sup> VDR, vitamin D receptor; 1 $\alpha$ ,25(OH)<sub>2</sub>D, 1 $\alpha$ ,25-dihydroxyvitamin D<sub>3</sub>.

One can derive at least 3 important conclusions, one prediction, and one expectation from the discoveries made over the past 30 y pertaining to the wide tissue distribution of both the VDR and the extrarenal 25(OH) $_2$ D $_3$ -1 $\alpha$ -hydroxylase

### Conclusions

1) Recent research has shown that vitamin D $_3$ 's biological sphere of influence is much broader than researchers originally thought, as shown by the tissue distribution of the VDR, from mediating only calcium homeostasis (intestine, bone, kidney, and parathyroid) to functioning as a pluripotent hormone in 5 physiologic arenas in which researchers have clearly identified additional biological actions of 1 $\alpha$ ,25(OH) $_2$ D $_3$  through the VDR. These physiologic arenas are the adaptive immune system, the innate immune system, insulin secretion by the pancreatic  $\beta$  cell, multifactorial heart functioning and blood pressure regulation, and brain and fetal development.

2) Researchers have also expanded the parent vitamin D $_3$ 's nutritional sphere of influence from a focus on bone health to include 5 additional physiologic systems.

3) The nutritional guidelines for vitamin D $_3$  intake must be carefully reevaluated to determine the adequate intake (balancing sunlight exposure with dietary intake) to achieve good health by involving all 36 target organs rather than just the first 4 target organs (intestine, kidney, bone, and parathyroid gland) that are considered for calcium homeostasis.

### Corollary

Given that vitamin D $_2$  is significantly less biologically active in humans than is vitamin D $_3$  (7), its biological use as a dietary supplement in the United States should be discontinued and its use in a high-dose form [eg, 500 000 IU/mL of ergocalciferol (vitamin D $_2$ )] in clinical studies (73, 74) and as described in the *Physicians Desk Reference* (75) should be replaced by a new formulation of high-dose vitamin D $_3$ .

### Prediction

Given the large expansion of the vitamin D endocrine system, the number of identified diseases or consequences of vitamin D deficiency or insufficiency will greatly increase to reflect the fact that the number of target organs for 1 $\alpha$ ,25(OH) $_2$ D $_3$  has increased  $\approx$ 9-fold since the discovery of the VDR in intestine, bone, kidney, and parathyroid tissues in the early 1970s.

### Expectation or hope

More than ever, we need to increase the amount of research on vitamin D (ie, increase funding from government agencies and pharmaceutical companies) to meet the challenge of maximizing the knowledge of how to use vitamin D in the context of the vitamin D endocrine system to preserve or improve the health of everyone on the planet.

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### REFERENCES

- Feldman D, Pike JW, Glorieux FH, eds. Vitamin D. San Diego, CA: Elsevier Academic Press, 2005.
- Norman AW, Bouillon R, Whiting SJ, Vieth R, Lips P. 13th Workshop

- consensus for vitamin D nutritional guidelines. *J Steroid Biochem Mol Biol* 2007;103:204–5.
- Gilchrest BA. Sun protection and vitamin D: three dimensions of obfuscation. *J Steroid Biochem Mol Biol* 2007;103:655–63.
- Wolpowitz D, Gilchrest BA. The vitamin D questions: how much do you need and how should you get it? *J Am Acad Dermatol* 2006;54:301–17.
- Standing Committee on the Scientific Evaluation of Dietary Reference Intakes, Food and Nutrition Board, Institute of Medicine. Vitamin D. Dietary reference intakes for calcium, magnesium, phosphorus, vitamin D, and fluoride. Washington, DC: National Academy Press, 1997.
- Trang H, Cole DE, Rubin LA, Pierratos A, Siu S, Vieth R. Evidence that vitamin D $_3$  increases serum 25-hydroxyvitamin D more efficiently than does vitamin D $_2$ . *Am J Clin Nutr* 1998;68:854–8.
- Armas LAG, Hollis BW, Heaney RP. Vitamin D $_2$  is much less effective than vitamin D $_3$  in humans. *J Clin Endocrinol Metab* 2004;89:5387–91.
- Bouillon R, Okamura WH, Norman AW. Structure-function relationships in the vitamin D endocrine system. *Endocr Rev* 1995;16:200–57.
- Horst RL, Reinhardt TA, Reddy GS. Vitamin D metabolism. In: Feldman D, Pike JW, Glorieux FH, eds. Vitamin D. San Diego, CA: Elsevier Academic Press 2005:15–36.
- Bishop JE, Collins ED, Okamura WH, Norman AW. Profile of ligand specificity of the vitamin D binding protein for 1 $\alpha$ ,25(OH) $_2$ -vitamin D $_3$  and its analogs. *J Bone Miner Res* 1994;9:1277–88.
- Henry HL. The 25-hydroxyvitamin D 1 $\alpha$  hydroxylase. In: Feldman D, Pike JW, Glorieux FH, eds. Vitamin D. San Diego, CA: Elsevier Academic Press, 2005:69–83.
- Barbour GL, Coburn JW, Slatopolsky E, Norman AW, Horst RL. Hypercalcemia in an anephric patient with sarcoidosis: evidence for extrarenal generation of 1,25-dihydroxyvitamin D. *N Engl J Med* 1981;305:440–3.
- Bises G, Kallay E, Weiland T, et al. 25-Hydroxyvitamin D $_3$ -1 $\alpha$ -hydroxylase expression in normal and malignant human colon. *J Histochem Cytochem* 2004;52:985–9.
- Adorini L, Penna G, Giarratana N, et al. Dendritic cells as key targets for immunomodulation by vitamin D receptor ligands. *J Steroid Biochem Mol Biol* 2004;89-90:437–41.
- Zehnder D, Bland R, Chana RS, et al. Synthesis of 1,25-dihydroxyvitamin D $_3$  by human endothelial cells is regulated by inflammatory cytokines: a novel autocrine determinant of vascular cell adhesion. *J Am Soc Nephrol* 2002;13:621–9.
- Eyles DW, Smith S, Kinobe R, Hewison M, McGrath JJ. Distribution of the vitamin D receptor and 1 $\alpha$ -hydroxylase in human brain. *J Chem Neuroanat* 2005;29:21–30.
- Segersten U, Holm PK, Bjorklund P, et al. 25-Hydroxyvitamin D $_3$  1 $\alpha$ -hydroxylase expression in breast cancer and use of non-1 $\alpha$ -hydroxylated vitamin D analogue. *Breast Cancer Res* 2005;7:R980–6.
- Townsend K, Evans KN, Campbell MJ, Colston KW, Adams JS, Hewison M. Biological actions of extra-renal 25-hydroxyvitamin D-1 $\alpha$ -hydroxylase and implications for chemoprevention and treatment. *J Steroid Biochem Mol Biol* 2005;97:103–9.
- Segersten U, Correa P, Hewison M, et al. 25-hydroxyvitamin D(3)-1 $\alpha$ -hydroxylase expression in normal and pathological parathyroid glands. *J Clin Endocrinol Metab* 2002;87:2967–72.
- Evans KN, Bulmer JN, Kilby MD, Hewison M. Vitamin D and placental-decidual function. *J Soc Gynecol Investig* 2004;11:263–71.
- Ma JF, Nonn L, Campbell MJ, Hewison M, Feldman D, Peehl DM. Mechanisms of decreased vitamin D 1 $\alpha$ -hydroxylase activity in prostate cancer cells. *Mol Cell Endocrinol* 2004;221:67–74.
- Bikle DD, Chang S, Crumrine D, et al. 25 Hydroxyvitamin D 1 $\alpha$  hydroxylase is required for optimal epidermal differentiation and permeability barrier homeostasis. *J Invest Dermatol* 2004;122:984–92.
- Huang DC, Papavasiliou V, Rhim JS, Horst RL, Kremer R. Targeted disruption of the 25-hydroxyvitamin D $_3$  1 $\alpha$ -hydroxylase gene in *ras*-transformed keratinocytes demonstrates that locally produced 1 $\alpha$ ,25-dihydroxyvitamin D $_3$  suppresses growth and induces differentiation in an autocrine fashion. *Mol Cancer Res* 2002;1:56–67.
- Moreno J, Krishnan AV, Feldman D. Molecular mechanisms mediating the anti-proliferative effects of vitamin D in prostate cancer. *J Steroid Biochem Mol Biol* 2005;97:31–6.
- Cross HS, Kallay E, Khorchide M, Lechner D. Regulation of extrarenal synthesis of 1,25-dihydroxyvitamin D $_3$  – relevance for colonic cancer prevention and therapy. *Mol Aspects Med* 2003;24:459–65.
- Eelen G, Gysemans C, Verlinden L, et al. Mechanism and potential of the

- growth-inhibitory actions of vitamin D and analogs. *Curr Med Chem* 2007;14:1893–910.
27. Norman AW, Mizwicki MT, Norman DPG. Steroid hormone rapid actions, membrane receptors and a conformational ensemble model. *Nat Rev Drug Discov* 2004;3:27–41.
  28. Whitfield GK, Jurutka PW, Haussler CA, et al. Nuclear vitamin D receptor: structure-function, molecular control of gene transcription and novel bioactions. In: Feldman D, Pike JW, Glorieux FH, eds. *Vitamin D*. San Diego, CA: Elsevier Academic Press 2005:219–328.
  29. Carlberg C. Current understanding of the function of the nuclear vitamin D receptor in response to its natural and synthetic ligands. *Recent Results Cancer Res* 2003;164:29–42.
  30. Huhtakangas JA, Olivera CJ, Bishop JE, Zanello LP, Norman AW. The vitamin D receptor is present in caveolae-enriched plasma membranes and binds 1 $\alpha$ ,25(OH)<sub>2</sub>-vitamin D<sub>3</sub> in vivo and in vitro. *Mol Endocrinol* 2004;18:2660–71.
  31. Razani B, Woodman SE, Lisanti MP. Caveolae: from cell biology to animal physiology. *Pharmacol Rev* 2002;54:431–67.
  32. Zanello LP, Norman AW. Rapid modulation of osteoblast ion channel responses by 1 $\alpha$ ,25(OH)<sub>2</sub>-vitamin D<sub>3</sub> requires the presence of a functional vitamin D nuclear receptor. *Proc Natl Acad Sci U S A* 2004;101:1589–94.
  33. Bula CM, Huhtakangas J, Olivera CJ, Bishop JE, Norman AW, Henry HL. Presence of a truncated form of the vitamin D receptor (VDR) in a strain of VDR-knockout mice. *Endocrinology* 2005;146:5581–6.
  34. Norman AW. Minireview: vitamin D receptor: new assignments for an already busy receptor. *Endocrinology* 2006;147:5542–8.
  35. Weaver CM, Fleet JC. Vitamin D requirements: current and future. *Am J Clin Nutr* 2004;80(suppl):1735S–9S.
  36. Hollis BW, Wagner CL. Normal serum vitamin D levels. *N Engl J Med* 2005;352:515–6.
  37. Kanis JA, Taylor CM, Douglas DL, Cundy T, Russell RGG. Effects of 24,25-dihydroxy-vitamin D<sub>3</sub> on its plasma level in man. *Metab Bone Dis Rel Res* 1981;3:155–8.
  38. Jastrup B, Moselilide L, Melsen F, Lund B, Lund BJ, Sorensen OG. Serum levels of vitamin D metabolites and bone remodelling in hyperthyroidism. *Metabolism* 1982;31:126–32.
  39. Pattanaungkul S, Riggs BL, Yergoy AL, Vieira NE, O'Fallon WM, Khosla S. Relationship of intestinal calcium absorption to 1,25-dihydroxyvitamin D [1,25(OH)<sub>2</sub>D] levels in young versus elderly women: evidence for age-related intestinal resistance to 1,25(OH)<sub>2</sub>D action. *J Clin Endocrinol Metab* 2000;85:4023–7.
  40. Muniz JF, Wehr CT, Wehr HM. Reverse phase liquid chromatographic determination of vitamins D<sub>2</sub> and D<sub>3</sub> in milk. *J Assoc Off Anal Chem* 1982;65:791–7.
  41. Hollis BW, Wagner CL, Drezner MK, Binkley NC. Circulating vitamin D<sub>3</sub> and 25-hydroxyvitamin D in humans: an important tool to define adequate nutritional vitamin D status. *J Steroid Biochem Mol Biol* 2007;103:631–4.
  42. Vieth R, Bischoff-Ferrari H, Boucher BJ, et al. The urgent need to recommend an intake of vitamin D that is effective. *Am J Clin Nutr* 2007;85:649–50.
  43. Need AG. Bone resorption markers in vitamin D insufficiency. *Clin Chim Acta* 2006;368:48–52.
  44. Fournier A, Fardellone P, Achard JM, et al. Importance of vitamin D repletion in uraemia. *Nephrol Dial Transplant* 1999;14:819–23.
  45. Lensmeyer GL, Wiebe DA, Binkley N, Drezner MK. HPLC method for 25-hydroxyvitamin D measurement: comparison with contemporary assays. *Clin Chem* 2006;52:1120–6.
  46. Hollis BW, Horst RL. The assessment of circulating 25(OH)D and 1,25(OH)<sub>2</sub>D: where we are and where we are going. *J Steroid Biochem Mol Biol* 2007;103:473–6.
  47. Saenger AK, Laha TJ, Bremner DE, Sadrzadeh SM. Quantification of serum 25-hydroxyvitamin D<sub>2</sub> and D<sub>3</sub> using HPLC-tandem mass spectrometry and examination of reference intervals for diagnosis of vitamin D deficiency. *Am J Clin Pathol* 2006;125:914–20.
  48. Holick MF. High prevalence of vitamin D inadequacy and implications for health. *Mayo Clin Proc* 2006;81:353–73.
  49. Nicolaysen R. XV. Studies upon the mode of action of vitamin D. III. The influence of vitamin D on the absorption of calcium and phosphorus in the rat. *Biochem J* 1937;31:122–9.
  50. Myrtle JF, Norman AW. Vitamin D: a cholecalciferol metabolite highly active in promoting intestinal calcium transport. *Science* 1971;171:78–82.
  51. Brickman AS, Coburn JW, Norman AW. Action of 1,25-dihydroxy-cholecalciferol, a potent, kidney-produced metabolite of vitamin D<sub>3</sub>, in uremic man. *N Engl J Med* 1972;287:891–5.
  52. Brickman AS, Coburn JW, Massry SG, Norman AW. 1,25-Dihydroxyvitamin D<sub>3</sub> in normal man and patients with renal failure. *Ann Intern Med* 1974;80:161–8.
  53. Suda T, Ueno Y, Fujii K, Shinki T. Vitamin D and bone. *J Cell Biochem* 2003;88:259–66.
  54. Mellanby E. Experimental rickets. Medical Research Council Special Report Series no. 61. London, United Kingdom: His Majesty's Stationery Office, 1921.
  55. McCollum EV, Simmonds N, Becker JE, Shipley PG. Studies on experimental rickets. XXI. An experimental demonstration of the existence of a vitamin which promotes calcium deposition. *J Biol Chem* 1922;53:293–312.
  56. Goldblatt H, Soames KN. A study of rats on a normal diet irradiated daily by the mercury vapor quartz lamp or kept in darkness. *Biochem J* 1923;17:294–7.
  57. Provvedini DM, Tsoukas CD, Deftos LJ, Manolagas SC. 1,25-dihydroxyvitamin D<sub>3</sub> receptors in human leukocytes. *Science* 1983;221:1181–3.
  58. Tsoukas CD, Provvedini DM, Manolagas SC. 1,25-dihydroxyvitamin D<sub>3</sub>: a novel immunoregulatory hormone. *Science* 1984;224:1438–40.
  59. van Etten EE, Mathieu C. Immunoregulation by 1,25-dihydroxyvitamin D<sub>3</sub>: basic concepts. *J Steroid Biochem Mol Biol* 2005;97:93–101.
  60. Wang TT, Nestel FP, Bourdeau V, et al. Cutting edge: 1,25-dihydroxyvitamin D<sub>3</sub> is a direct inducer of antimicrobial peptide gene expression. *J Immunol* 2004;173:2909–12.
  61. Liu PT, Stenger S, Li H, et al. Toll-like receptor triggering of a vitamin D-mediated human antimicrobial response. *Science* 2006;311:1770–3.
  62. Schaubert J, Dorschner RA, Coda AB, et al. Injury enhances TLR2 function and antimicrobial peptide expression through a vitamin D-dependent mechanism. *J Clin Invest* 2007;117:803–11.
  63. Norman AW, Frankel BJ, Heldt AM, Grodsky GM. Vitamin D deficiency inhibits pancreatic secretion of insulin. *Science* 1980;209:823–5.
  64. Kadowaki S, Norman AW. Demonstration that the vitamin D metabolite 1,25(OH)<sub>2</sub>-vitamin D<sub>3</sub> and not 24R,25(OH)<sub>2</sub>-vitamin D<sub>3</sub> is essential for normal insulin secretion in the perfused rat pancreas. *Diabetes* 1985;34:315–20.
  65. Mathieu C, Badenhoop K. Vitamin D and type 1 diabetes mellitus: state of the art. *Trends Endocrinol Metab* 2005;16:261–6.
  66. Giulietti A, Gysemans C, Stoffels K, et al. Vitamin D deficiency in early life accelerates type 1 diabetes in non-obese diabetic mice. *Diabetologia* 2004;47:451–62.
  67. Chiu KC, Chu A, Go VL, Saad MF. Hypovitaminosis D is associated with insulin resistance and beta cell dysfunction. *Am J Clin Nutr* 2004;79:820–5.
  68. Harms LR, Eyles DW, McGrath JJ, et al. Developmental vitamin D deficiency alters adult behaviour in 129/SvJ and C57BL/6J mice. *Behav Brain Res* 2008;187:343–50.
  69. Burne TH, Feron F, Brown J, Eyles DW, McGrath JJ, Mackay-Sim A. Combined prenatal and chronic postnatal vitamin D deficiency in rats impairs prepulse inhibition of acoustic startle. *Physiol Behav* 2004;81:651–5.
  70. Simpson RU, Hershey SH, Nibbelink KA. Characterization of heart size and blood pressure in the vitamin D receptor knockout mouse. *J Steroid Biochem Mol Biol* 2007;103:521–4.
  71. Maiya S, Sullivan I, Allgrove J, et al. Hypocalcaemia and vitamin D deficiency: an important, but preventable cause of life threatening infant heart failure. *Heart* 2008;94:581–4.
  72. Li YC, Kong J, Wei M, Chen ZF, Liu SQ, Cao LP. 1,25-Dihydroxyvitamin D<sub>3</sub> is a negative endocrine regulator of the renin-angiotensin system. *J Clin Invest* 2002;110:229–38.
  73. Boyle MP, Noschese ML, Watts SL, et al. Failure of high-dose ergocalciferol to correct vitamin D deficiency in adults with cystic fibrosis. *Am J Respir Crit Care Med* 2005;172:212–7.
  74. Munns C, Zacharin MR, Rodda C, et al. Prevention and treatment of infant and childhood vitamin D deficiency in Australia and New Zealand: a consensus statement. *Med J Aust* 2006;185:268–72.
  75. Physicians desk reference. Montvale, NJ: Medical Economics Company, 2002.